Acta Crystallographica Section C Crystal Structure Communications

ISSN 0108-2701

6-Deoxyjacareubin

Antonio C. Doriguetto,^a* Marcelo H. Santos,^b Javier A. Ellena^a and Tanus J. Nagem^c

^aInstituto de Física de São Carlos, Universidade de São Paulo, Caixa Postal 369 – CEP 13560-970, São Carlos, SP, Brazil, ^bDepartamento de Química, Universidade Federal de Minas Gerais, CP 702, 31270-901 Belo Horizonte, MG, Brazil, and ^cDepartamento de Química, Universidade Federal de Ouro Preto, CEP 35400-000 Ouro Preto, MG, Brazil Correspondence e-mail: dorigue@if.sc.usp.br

Received 22 March 2001 Accepted 4 June 2001

The natural compound 5,10-dihydroxy-2,2-dimethylpyrano-[3,2-b]xanthen-6(2*H*)-one (6-deoxyjacareubin), C₁₈H₁₄O₅, was isolated from leaves of *Vismia latifolia* (Guttiferae family). The compound has four six-membered rings. The molecule has two planar benzenoid and one planar pyranoid ring, plus a pyranoid ring in a distorted chair conformation. The crystal is stabilized by one intra- and one intermolecular hydrogen bond.

Comment

The title compound, (I), is a natural xanthone. Xanthones are heterocyclic ring systems with many pharmacological applications (Santos, Nagem, Silva & da Silva, 2000). It was isolated from *Vismia latifolia* Choisy (Syn. *Hypericum latifolium* Aubl.). This species belongs to the Guttiferae family, subfamily Hypericoideae and tribe *Vismieae* and is a tree commonly known in the Bahia state (Brazil) as pau-de-sangue. This plant is widely used in traditional medicine as a tonic and febrifugal agent (Corrêa, 1978). Previous works have reported the presence of several xanthones, terpenoids, benzophenones, flavonoids and anthranoids in *Vismia* species (Santos *et al.*, 1999; Santos, Nagem, da Silva & Silva, 2000; Nagem & De Oliveira, 1997; Peres & Nagem, 1997; Nagem & Alves, 1995; Nagem & Ferreira, 1993; Delle Monache *et al.*, 1983).

As part of a chemotaxonomic study of the Guttiferae family (*Vismia* genus), we have determined the structure of the compound known as 6-deoxyjacareubin, (I). We have identified (I) by spectroscopic methods (UV, EI–MS, and ¹H and ¹³C NMR) and the presence of the γ -pyrone was confirmed by the X-ray data. This compound was first isolated from *Calophyllum inophyllum* by Jackson *et al.* (1969). It is described as an antifungal agent against *Postia placenta* (Reyes Chilpa *et al.*, 1997) and *Cladosporium cucumerinum* (Rocha *et al.*, 1994). For this reason, we have also tested its biological activity against fungi [*Cândida albicans, Aspergillus ochraceus* (*allutaceus*) and *Penicillium citrinum*]

and bacteria (*Escherichia coli* and *Streptococcus muttans*). Our tests, however, did not show biological inhibition on these organisms.



Fig. 1 is an ORTEP-3 (Farrugia, 1997) view of the title compound. As expected, the molecule is almost flat. All atoms in the A, B and C rings lie within 0.149 (1) Å of the leastsquares plane through the three-ring system. The A, B and C rings are also individually almost planar, including the O1, O2 and O3 atoms linked to them. The largest deviations from the individual least-squares planes are 0.006 (2), 0.037 (1) and 0.009 (1) Å for rings A, B and C, respectively. The leastsquares planes of the A and B rings form an angle of 3.64 $(8)^{\circ}$, those of the B and C rings form an angle of 3.69 (8)°, and those of the A and C rings form an angle of 7.31 (9)°. The planes of the A and C ring systems intersect on a line which is approximately through the middle of the B ring. A folding point along an imaginary line drawn between O5 and C6 defines an angle of 6.55 (6)° (least-squares planes of rings A and C calculated by including O5 and C6). Ring D is in a deformed chair conformation. The weighted average absolute torsion angle (Domenicano *et al.*, 1975) in the D ring is 27 (8)°. The main bond lengths and angles are given in Table 1. The mean value of the intramolecular C-C bond distances within the A and C benzenoid rings is 1.39(1) Å, which agrees well with the normal aromatic value. In the same way, the mean bond angles in rings A and C are $120 (2)^{\circ}$. The average value of the two C-O5 bond lengths in pyranoid ring B is 1.373 (2) Å. The observed geometry of pyranoid ring B agrees well with similar pyranxanthone geometries (Ho et al., 1987; Vijavalakshmi et al., 1987; Ferguson et al., 1985; Söderholm et al., 1976).

The title molecule exhibits a moderated intramolecular hydrogen bond, $O3-H3'\cdots O2$, with an $O\cdots O$ distance of 2.561 (2) Å (Table 2). The intermolecular hydrogen bond



Figure 1

ORTEP-3 (Farrugia, 1997) view of 6-deroxyjacareubin, showing the atom and ring labelling and 50% probability ellipsoids.

between hydroxyl group O1-H1' and adjacent carboxyl oxygen O2 at (x + 1, y, z) [the O···O distance is 2.778 (2) Å] stabilizes the structure and gives rise to a chain parallel to the [100] direction (Fig. 2). The separation between the least-squares planes through the molecules at (x, y, z) and (-x, -y, -z) is 3.67 (2) Å.



Figure 2

View of the chain of hydrogen bonds parallel to [100]. Symmetry codes from top to bottom are (1 + x, y, z), (x, y, z) and (1 - x, y, z).

Experimental

Dried and ground leaves (1502.0 g) of *Vismia latifolia* were extracted (at room temperature) with *n*-hexane (32.85 g) and EtOH (200.0 g) in succession. The ethanol extract was stirred with water for 24 h and filtered. The soluble portion was extracted with CHCl₃, EtOAc and *n*-butanol in succession. The solid obtained by evaporation (under vacuum) of the fraction soluble in CHCl₃ (30.0 g) was chromato-graphed on silica-gel (Merck, 352 g) column chromatography and eluted with CH₂Cl₂, EtOAc and EtOH. The 90 fractions obtained yielded nine groups (D1–D9). D3 (fractions 4–6, 658.0 mg) was washed with petrol ether giving an insoluble portion that was washed with methanol yielding 6-deoxyjacareubin (101.0 mg) as a yellow solid. The powder mass obtained was crystallized from methanol by slow evaporation at room temperature.

Crystal data

$C_{18}H_{14}O_5$	Z = 2
$M_r = 310.29$	$D_x = 1.424 \text{ Mg m}^{-3}$
Triclinic, $P\overline{1}$	Mo $K\alpha$ radiation
a = 7.4450 (2) Å	Cell parameters from 3315
b = 9.5200(3) Å	reflections
c = 10.9390(3) Å	$\theta = 1.0-27.5^{\circ}$
$\alpha = 105.051 \ (2)^{\circ}$	$\mu = 0.11 \text{ mm}^{-1}$
$\beta = 103.154 \ (2)^{\circ}$	T = 293 (2) K
$\gamma = 93.083 \ (2)^{\circ}$	Prism, yellow
$V = 723.80 (4) \text{ Å}^3$	$0.24 \times 0.17 \times 0.12 \text{ mm}$

Table 1

Selected geometric parameters (Å, °).

01-C1	1.354 (2)	C10-C11	1.327 (3)
O3-C8	1.346 (2)	C11-C12	1.504 (3)
O4-C15	1.360 (2)	C12-C13	1.517 (2)
O4-C12	1.470 (2)	C12-C14	1.522 (3)
C9-C10	1.458 (2)		
O4-C12-C11	109.3 (1)	O4-C12-C14	107.4 (2)
O4-C12-C13	104.5 (2)	C11-C12-C14	111.8 (2)
C11-C12-C13	112.2 (2)	C13-C12-C14	111.3 (2)

Table 2

Hydrogen-bonding geometry (Å, °).

$D - H \cdots A$	D-H	$H \cdot \cdot \cdot A$	$D{\cdots}A$	$D - H \cdots A$
$\begin{array}{c} O1 - H1' \cdots O2^{i} \\ O3 - H3' \cdots O2 \end{array}$	0.87 (3) 0.89 (3)	1.97 (3) 1.74 (3)	2.778 (2) 2.561 (2)	154 (3) 152 (4)

Symmetry code: (i) 1 + x, y, z.

Data collection

Nonius KappaCCD diffractometer φ and ω scans with κ offsets 6210 measured reflections 3319 independent reflections 2674 reflections with $I > 2\sigma(I)$	$R_{\text{int}} = 0.014$ $\theta_{\text{max}} = 27.5^{\circ}$ $h = 0 \rightarrow 9$ $k = -12 \rightarrow 12$ $l = -14 \rightarrow 13$
Refinement	
Refinement on F^2 P(E) = 0.051	$w = 1/[\sigma^2(F_o^2) + (0.0624P)^2 + 0.4000P]$
$wR(F^2) = 0.051$ $wR(F^2) = 0.150$	where $P = (F_o^2 + 2F_c^2)/3$
S = 1.02	$(\Delta/\sigma)_{\rm max} = 0.003$
3319 reflections	$\Delta \rho_{\rm max} = 0.41 \ {\rm e} \ {\rm \AA}^{-3}$
218 parameters	$\Delta \rho_{\rm min} = -0.49 \ {\rm e} \ {\rm \AA}^{-3}$
H atoms treated by a mixture of	,
independent and constrained	
refinement	

The H atoms of the phenyl and methyl groups were positioned stereochemically and were refined with fixed individual displacement parameters $[U_{iso}(H) = 1.2U_{eq}(C) \text{ or } 1.5U_{eq}(C_{methoxy})]$ using a riding model with aromatic C-H = 0.93 Å and methyl C-H = 0.98 Å. The hydroxyl H atoms were located by difference Fourier synthesis and were set as isotropic.

Data collection: *COLLECT* (Nonius, 1997–2000); cell refinement: *HKL SCALEPACK* (Otwinowski & Minor, 1997); data reduction: *HKL DENZO* and *SCALEPACK* (Otwinowski & Minor, 1997); program(s) used to solve structure: *SHELXS86* (Sheldrick, 1985); program(s) used to refine structure: *SHELXL97* (Sheldrick, 1997); molecular graphics: *ORTEP-3 for Windows* (Farrugia, 1997); software used to prepare material for publication: *WinGX* (Farrugia, 1999).

This work was supported by Brazilian agencies FAPESP (Projeto Temático No. 98/12151–1) and FAPEMIG. MHS is grateful to CAPES and CNPq for providing a doctoral fellowship. JE and ACD thank FAPESP and CNPq, respectively, for postdoctoral fellowships.

Supplementary data for this paper are available from the IUCr electronic archives (Reference: DA1189). Services for accessing these data are described at the back of the journal.

References

- Corrêa, M. P. (1978). Dicionário das Plantas Uteis do Brasil e das Plantas Exóticas Cultivadas. Rio de Janeiro: Imprensa Nacional.
- Delle Monache, F., Mac-Quhae, M. M., Delle Monache, G., Marini-Bettolo, G. B. & Alves De Lima, R. (1983). *Phytochemistry*, 22, 227–232.
- Domenicano, A., Vaciago, A. & Coulson, C. A. (1975). Acta Cryst. B31, 221-234.
- Farrugia, L. J. (1997). J. Appl. Cryst. 30, 565.
- Farrugia, L. J. (1999). J. Appl. Cryst. 32, 837-838.
- Ferguson, G., Kaitner, B., Gilmore, J., Omuaru, V. O. T. & Whalley, W. B. (1985). J. Chem. Soc. Perkin Trans. 1, 7, 1343–1347.
- Ho, D. K., McKenzie, A. M., Byrn, S. R. & Cassady, J. M. (1987). *J. Org. Chem.* **52**, 342–347.
- Jackson, B., Locksley, H. D. & Scheinma, F. (1969). Phytochemistry, 8, 927-929.
- Nagem, T. J. & Alves, V. L. (1995). Fitoterapia, 66, 278.
- Nagem, T. J. & de Oliveira, F. F. J. (1997). Braz. Chem. Soc. 8, 505-508.
- Nagem, T. J. & Ferreira, M. A. (1993). Fitoterapia, 64, 382-383.

- Nonius (1998). COLLECT. Nonius BV, Delft, The Netherlands.
- Otwinowski, Z. & Minor, W. (1997). Methods Enzymol. 276, 307-326.
- Peres, V. & Nagem, T. J. (1997). Phytochemistry, 44, 191-214.
- Reyes Chilpa, R., Jimenez Estrada, M. & Estrada Muniz, E. (1997). J. Chem. Ecol. 23, 1901–1911.
- Rocha, L., Marston, A., Kaplan, M. A. C., Stoeckli-Evans, H., Thull, U., Testa, B. & Hostettmann, K. (1994). *Phytochemistry*, **36**, 1381–1385.
- Santos, M. H., Nagem, T. J., da Silva, M. C. & Silva, L. G. F. (2000). J. Braz. Chem. Soc. 11, 537–539.
- Santos, M. H., Nagem, T. J., de Oliveira, T. T. & Braz, R. (1999). *Quím. Nova*, **22**, 654–660.
- Santos, M. H., Nagem, T. J., Silva, L. G. F. E. & da Silva, M. C. (2000). Magnet. Reson. Chem. 38, 1027–1030.
- Sheldrick, G. M. (1985). SHELXS86. University of Göttingen, Germany.
- Sheldrick, G. M. (1997). SHELXL97. University of Göttingen, Germany.
- Söderholm, M., Sonnerstam, U., Norrestam, R. & Palm, T.-B. (1976). Acta Cryst. B32, 3013–3018.
- Vijayalakshmi, J., Rajam, S. S. & Srinivasan, R. (1987). Acta Cryst. C43, 2108–2110.